

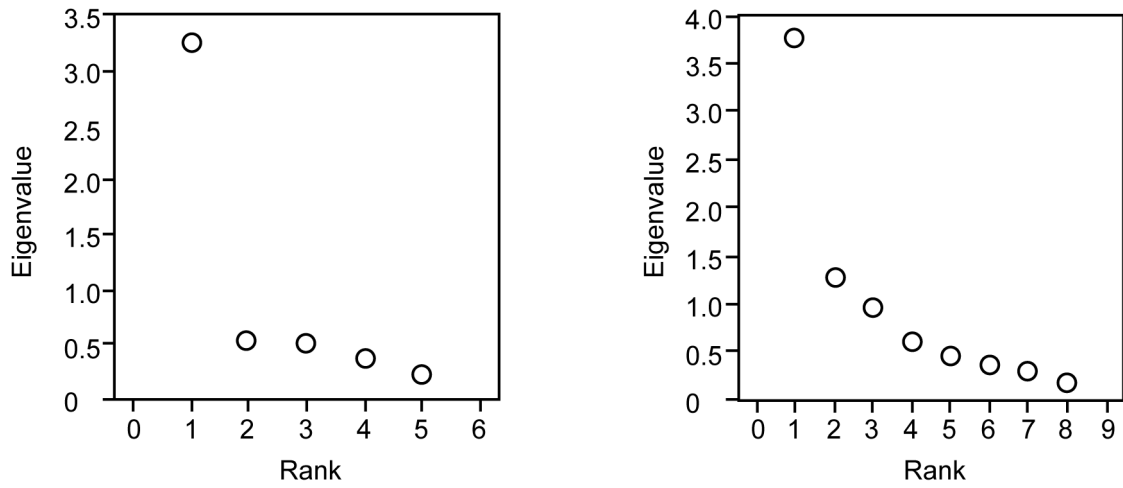
1 **Electronic Supplementary Material: Reader, Hager & Laland**

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Supplemental Data: Figures

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6 Figure S1. Scree plots for principal components analyses. The plots illustrate the
7 dominance of the first component. Left panel: 5-variable analysis. Right panel: 8-variable
8 analysis.

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Supplemental Data: Tables

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Table S1. Principal component and factor analyses. Non-rotated and oblique rotated solutions gave broadly similar results to the PCA presented in the main text, as did factor analyses.

(a) 5-variable analyses. All analyses extract one component/factor.

	PCA	FA
<i>Extractive foraging</i>	.84	.65
<i>Innovation</i>	.75	.47
<i>Social learning</i>	.82	.60
<i>Tactical deception</i>	.74	.48
<i>Tool use</i>	.88	.75
VARIANCE	65%	58%

(b) 8-variable analyses. All analyses extract two components/factors

Factor	PCA (non-rotated)		PCA (oblique)		FA (non-rotated)	
	1	2	1	2	1	2
<i>Diet breadth</i>	.57	.53	.21	.69	.53	.48
<i>Extractive foraging</i>	.85	-.24	.88	.00	.84	-.20
<i>Innovation</i>	.72	-.31	.81	-.10	.68	-.26
<i>Percent fruit</i>	.39	.58	.02	.70	.30	.45
<i>Population group size</i>	.23	.65	-.16	.72	.19	.32
<i>Social learning</i>	.85	-.05	.78	.20	.83	.01
<i>Tactical deception</i>	.76	.06	.64	.28	.70	.09
<i>Tool use</i>	.83	-.30	.90	-.07	.81	-.23
VARIANCE	47%	16%	47%	16%	43%	9%
INTERCORRELATION			.28			

20 Table S2. Regressions of brain size on g_1 with the effect of body mass removed. The
 21 effect of body mass was statistically controlled for through multiple regression, for both
 22 across-species and independent contrast analyses. The relationships with brain measures
 23 1-3 are weaker than when body mass is not included, with some non-significant (see
 24 Table 4 main body), reflecting the well-known finding that ratio and absolute brain
 25 measures are to some degree confounded with body size [1]. Deaner *et al.* [2] have
 26 argued that absolute brain measures correlate more strongly with cognitive performance
 27 in primates than do relative brain measures. The observation that the relationship
 28 between primate g and brain size measures remain significant, or near significant, for the
 29 independent contrast data suggests that the relationship is not solely a confound of body
 30 mass.

31

<i>Brain measure</i>	<i>Analysis</i>	<i>N</i>	<i>t</i>	<i>P</i>
1. Neocortex ratio	Across-species	29	1.71	0.10
	Independent contrasts	28	2.79	0.01
2. Executive brain ratio	Across-species	20	2.63	0.02
	Independent contrasts	19	1.94	0.07
3. Ln (neocortex volume)	Across-species	29	1.32	0.20
	Independent contrasts	28	2.08	0.05
4. Residuals of neocortex on rest of brain	Across-species	29	0.37	0.72
	Independent contrasts	28	1.13	0.27

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33

33 Table S3. Group size remains a significant predictor of brain size for two brain measures.
 34 Since there are statistically significant positive relationships between population group
 35 size and both tactical deception and diet breadth (Table 3, main text), we conducted
 36 multiple regressions to investigate whether group size remained a predictor of brain size
 37 when deception and diet breadth were included as independent variables. Group size
 38 remained a significant predictor of brain size for two brain measures.

<i>Brain measure</i>	<i>Analysis</i>	<i>N</i>	<i>t</i>	<i>P</i>
1. Neocortex ratio	Across-species	30	3.75	0.0009
	Independent contrasts	29	2.89	0.008
2. Executive brain ratio	Across-species	20	1.19	0.25
	Independent contrasts	19	0.33	0.74
3. Ln (neocortex size)	Across-species	30	2.00	0.06
	Independent contrasts	29	0.20	0.84
4. Residuals of neocortex on rest of brain	Across-species	30	3.36	0.002
	Independent contrasts	29	2.32	0.03

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41 **Supplemental Experimental Procedures and Results**

42

43 **1. Data collection**

44 (a) *Innovation, social learning, tool use, extractive foraging*. We surveyed articles in
45 four journals (*Primates, American Journal of Primatology, Folia Primatologica* and the
46 *International Journal of Primatology*), as well as other relevant literature, collating
47 examples of behaviour in each category. These publications were chosen because they
48 are the leading primate behaviour journals, and collectively publish the bulk of primate
49 behavioural research. Following previous work [3-5], we used keywords (e.g. ‘novel’,
50 ‘extract’, or ‘traditional’) to classify behaviour patterns (e.g. as ‘innovation’, ‘extractive
51 foraging’ or ‘social learning’), so that the judgment of whether a behaviour pattern
52 qualified in a category was made by the expert author of the article. This approach aims
53 to avoid subjective bias on our part imposed during data collation. The surveyed articles
54 were published between 1925-2000, with considerable change in taxonomic
55 nomenclature during this period. Where species have had more than one name, or have
56 experienced changes or ambiguities in nomenclature, we searched and categorized using
57 relevant alternative names, and favoured grouping over splitting species (Table E2).
58 Species names were subsequently checked against the leading contemporary taxonomy
59 [6]. Further details on data collection methods, and examples of behaviour classified as
60 innovations are described elsewhere [3,7-9].

61 Extractive foraging is defined as feeding on foods that must first be extracted
62 from matrices in which they are embedded or encased, including nutmeat, shellfish,
63 snails, eggs, brains, bone marrow, roots, tubers, and ant and termite mounds [10].
64 Extractive foraging data were validated against an existing categorization [10] used in

65 previous work on cognitive evolution [11]. There were significantly more reports of
66 extractive foraging in genera that Gibson [10] categorized as extractive foragers (*Ateles*,
67 *Callimico*, *Callithrix*, *Cebuella* [Gibson specifies only 'marmosets'], *Cebus*,
68 *Daubentonia*, *Gorilla*, *Lagothrix*, *Macaca*, *Miopithecus*, *Papio*, *Pan*, *Pongo*, *Saimiri*),
69 compared with Gibson's non-extractive foragers (Mann-Whitney test: $U = 131.5$, $n_1 = 14$,
70 $n_2 = 44$, $p = 0.001$), a result that holds when prosimians and tarsiers are excluded from the
71 analysis (Mann-Whitney: $U = 73.0$, $n_1 = 13$, $n_2 = 22$, $p = 0.02$) or when based on reports
72 per unit of research effort (all primates: Mann-Whitney: $U = 141.0$, $n_1 = 14$, $n_2 = 44$, $p =$
73 0.002 ; simians: Mann-Whitney: $U = 79.0$, $n_1 = 13$, $n_2 = 22$, $p = 0.03$). Thus although
74 concern has been expressed that quantitative measures of extractive foraging are difficult
75 to assemble [12], independently compiled classification schemes corroborate our
76 extractive foraging measure. Moreover, experimental studies of extractive foraging in
77 callitrichid monkeys show that published reports of extractive foraging predict
78 performance in response to novel extractive foraging tasks under experimental conditions
79 [13], supporting the use of literature reports to estimate species differences in extractive
80 foraging.

81 The frequencies of social learning, innovation, tool use, extractive foraging, and
82 tactical deception were corrected for research effort by taking the perpendicular offset
83 residuals from a linear regression of research effort (see below) on observation frequency
84 (both log transformed), forced through the origin. Linear regression is suitable because a
85 clear direction of causality is established [14], with orthogonal offsets appropriate
86 because there is measurement error in both dependent and independent variable. While
87 we correct for research effort we cannot rule out the possibility of systematic reporting
88 biases distorting our findings. However, below we describe measures that we adopted

89 (e.g. repeating the analyses with great apes removed) which found no evidence that such
90 biases were operating. Reader & MacDonald [8,9] further discuss the methodology of
91 accounting for differences in research effort.

92

93 (b) *Tactical deception*. Deception data were taken from Byrne & Whiten [15], who
94 compiled instances of tactical deception by surveying primatologists using a standardized
95 request. While this survey has drawn criticism (see peer commentary to [16], it has been
96 defended [16], and has the advantage of being largely independent of other datasets. We
97 counted observations of tactical deception categorized by Byrne & Whiten [15] as level 1
98 ('evidence for tactical deception outweighs competing explanations') and above, with
99 repeated observations of tactical deception in the same category and by the same
100 observer considered as a single datum. To account for the fact that some instances of
101 tactical deception may qualify for other measures of behavioural flexibility (e.g.
102 innovative deception; [17], we noted acts simultaneously classified as deception and
103 another measure during our data collection and removed these from the database.
104 Similarly, we removed from our database examples of 'social tool use' [18], which we
105 did not consider as tool use, despite some arguments to the contrary [18].

106

107 (c) *Diet breadth*. Diet breadth data were compiled by allocating prey to 13 categories (1.
108 Invertebrate prey, 2. Vertebrate prey, 3. Fruit, fungus, and honey, 4. Seeds, nuts, 5.
109 Exudates, 6. Flowers, 7. Nectar/Pollen, 8. Roots, tubers, bulbs, truffles, 9. Leaves, shoots,
110 stems, herbs, buds, 10. Wood, bamboo, 11. Bark, 12. Pith, and 13. Lichen), with each
111 species given a score between 1-13 to specify diet breadth. Dietary items consumed were
112 taken from a single source [19], with subspecies' diets pooled. Initial categorization was

113 based on the consensus of categories utilized in previous studies [20-25], with additional
114 categories to account for additional foodstuffs. These categories were then collapsed to
115 the 13 final categories based on consideration of foraging strategy, covariance between
116 diet types, nutritional content, and the contribution of a particular foodstuff to the diet.
117 For example, all 11 fungus-eating species eat fruit, and because fungi are likely to be
118 spatially and temporally distributed in a similar manner to fruit, these two categories
119 were combined. This new dietary breadth measure is validated by strong correlations
120 with previously published dietary breadth estimates (versus Jernvall & Wright [21],
121 whose data were also compiled from Rowe [19]: $r^2 = 0.58$, $F_{1,209} = 287.04$, $p < 0.0001$;
122 versus Eeley & Foley [20], $r^2 = 0.46$, $F_{1,41} = 34.36$, $p < 0.0001$).

123

124 *(d) Percent fruit (and seeds) in diet.* Percentage fruit in diet data were compiled from
125 Smuts *et al.* [26], and percentage fruit and seeds from Kaplan & Robson [27]. The
126 former measure has the advantage that the rapid change in state (e.g. ripening) and spatial
127 distribution of fruits, but not seeds, lends itself well to notions of ecological intelligence
128 [28]. However, this first dataset includes only 52 species. The Kaplan & Robson [27]
129 measure compensates for the inclusion of seeds by providing data for substantially more
130 species (104). Accordingly here our analyses utilize the latter measure [27]. Values for
131 species common to the Kaplan & Robson [27] and Smuts *et al.* [26] datasets differ by
132 less than 5% [27].

133

134 *(e) Grooming clique size, population group size and foraging group size.* Data on
135 grooming clique size, so-called population group size (social group size), and foraging
136 group size were obtained from published sources [29-31]. Kudo & Dunbar [29] argue

137 that grooming clique size is the best population-size measure of the cognitive demands of
138 tracking social relationships, but this measure is available for only 29 species, reducing
139 the power and generality of analyses. Conversely, foraging group size is probably not a
140 good measure of such cognitive demands [32]. Accordingly, our analyses here of group
141 size use population group size, for which there are data for 169 species. All three group
142 size measures co-vary strongly.

143

144 (f) *Research effort*. The intensity of behavioural research on each species was assessed by
145 surveying the number of published articles per species in the *Zoological Record*, between
146 years 1993-2001. This source was suitable since it includes primate behavioural research
147 but not biomedical studies. We also collated data for species not in the behavioural
148 flexibility databases, to allow genus-level estimates of research effort to be made. Five
149 additional measures of research effort were computed by searching the number of
150 published articles on each species in five leading behaviour journals (*Primates*,
151 *International Journal of Primatology*, *American Journal of Primatology*, *Folia*
152 *Primatologica*, *Animal Behaviour*) by searching www.scopus.com, 1960-2005, for a
153 given primate species, using all common alternative Latin names, in articles, abstracts or
154 keywords of all document types.

155

156 (g) *Body mass*. Body mass data were taken from Smith & Cheverud [33], supplemented
157 by data from additional sources [34-36] that met the criteria of being based on
158 measurements of > 2 individuals of each sex, or > 8 unsexed individuals.

159

160 (h) *Brain component volumes*. Volumes of relevant brain regions were taken from
161 Stephan *et al.* [37], supplemented by additional data from both serial sections of brains
162 and magnetic resonance imaging [8,38-43]. The compatibility of utilizing these separate
163 sources was validated by comparing measures on the 8 species for which both slices and
164 scan methods had been employed, which exhibited no significant differences (paired t-
165 test, $t = 0.53$, $df = 7$, $p = 0.61$), a finding consistent with other studies comparing
166 alternative methods for measuring primate brains [39]. In total, brain volumes for 56
167 primate species were used (Table E2).

168

169 (i) *Cognitive performance in the laboratory*. Deaner *et al.* [44] compiled a continuous
170 index of global variable means for the relative performance of 24 primate genera in
171 laboratory tests of cognition. The tests are a heterogeneous complex of tasks, including
172 detour problems, patterned string problems, tool use, invisible displacement tasks, object
173 discrimination learning sets, reversal learning, oddity learning, delayed response tasks,
174 and sorting tasks. We examined to what extent Deaner *et al.*'s index (reduced model) can
175 be predicted by our primate g measure, using linear regression. Genus-level analyses
176 were based on the genera named in the composite phylogeny used for phylogenetic
177 analysis [45] (see below). However, a slightly modified classification scheme was used
178 for analyses including the Deaner *et al.* index, to match their listed genera: *Cebuella* was
179 grouped with *Callithrix*; *Mirza* with *Microcebus*; *Petterus* was renamed *Eulemur*; and
180 *Euoticus*, *Galago*, *Galagoides* and *Otolemur* were grouped together under *Galago*. Two
181 further measures of primate learning performance in the laboratory were also employed,
182 based on Riddell & Corl [46]. These authors compiled published performance data in six
183 different learning tasks. The largest single data set ('learning sets') includes data on six

184 species of primate (excluding humans), and by comparing performance across the
 185 various tasks it was possible to rank nine primate species ('combined Riddell rank'; [8].
 186 In ascending rank the species are *Callithrix jacchus* and *Saimiri sciureus* (ranked
 187 equally), *Papio hamadryas*, *Galago senegalensis*, *Cebus albifrons*, *Ateles geoffroyi*,
 188 *Macaca mulatta*, *Cebus capucinus*, and *Pan troglodytes*. Spearman rank correlation was
 189 employed to examine the relationship between ranked primate *g* and laboratory
 190 performance.

191

192 (j) *Summary*. Table E1 summarizes the number of records and number of species covered
 193 by each variable. Studies covered 251 species of primates, but because of overlapping
 194 datasets reported analyses range from 8 to 62 species. Table E2 lists the names and
 195 alternative Latin names of (i) those species used in analyses, (ii) those species for which
 196 brain data were available, and (iii) those species with non-zero scores in at least one of
 197 the five behavioural flexibility measures.

198

199 Table E1. Details of the number of records and number of species covered by each
 200 variable.

<i>Measure</i>	<i>Number of records</i>	<i>Number of species</i>
Innovation	588	62
Social learning	469	62
Tool use	656	62
Extractive foraging	430	62
Tactical deception	95	62
Diet breadth	N/A	236
% fruit and seeds [27]	N/A	104
Population group size [30]	N/A	169
Laboratory performance [46]	N/A	8
Neocortex ratio	N/A	56
Executive brain ratio	N/A	56
Ln (neocortex volume)	N/A	56
Residuals of neocortex on rest of brain	N/A	56

201

202

203 Table E2. List of species names and alternative names used to group the same species
 204 and in surveys of research effort, those species for which brain data were available, and
 205 those species with non-zero scores in at least one of the five behavioural flexibility
 206 measures (Y= data available). The initial list of species names came from the sources
 207 employed in our analyses [8,9,19,37,45,47], and were subsequently checked against the
 208 leading contemporary taxonomy [6]. Thus the table does not contain all current primates
 209 or all alternative names, and includes misspellings present in published literature.

Species	Alternative names	Brain data available	Behavioural data available
<i>Allenopithecus nigroviridis</i>			
<i>Allocebus trichotis</i>			
<i>Alouatta belzebul</i>			
<i>Alouatta caraya</i>			
<i>Alouatta coibensis</i>	<i>A. palliata coibensis</i>		
<i>Alouatta fusca</i>	<i>Alouatta guariba</i>		Y
<i>Alouatta palliata</i>		Y	Y
<i>Alouatta pigra</i>	<i>A. palliata pigra</i>		
<i>Alouatta sara</i>	<i>A. seniculus sara</i>		
<i>Alouatta seniculus</i>		Y	Y
<i>Aotus azarai</i>	<i>Aotus azarae</i>		
<i>Aotus brumbacki</i>			
<i>Aotus hershkovitzi</i>			
<i>Aotus infulatus</i>			
<i>Aotus lemurinus</i>			
<i>Aotus miconax</i>			
<i>Aotus nancymae</i>			
<i>Aotus nigriceps</i>			
<i>Aotus trivirgatus</i>		Y	
<i>Aotus vociferans</i>			Y
<i>Arctocebus calabarensis</i>			
<i>Ateles belzebuth</i>			
<i>Ateles fusciceps</i>			
<i>Ateles geoffroyi</i>		Y	Y
<i>Ateles paniscus</i>			
<i>Avahi laniger</i>	<i>Avahi laniger laniger</i>	Y	
<i>Avahi occidentalis</i>	<i>Avahi laniger occidentalis</i>	Y	
<i>Brachyteles arachnoides</i>			
<i>Cacajao calvus</i>			
<i>Cacajao melanocephalus</i>			
<i>Cacajao rubicundus</i>	<i>Cacajao calvus rubicundus</i>		
<i>Callicebus brunneus</i>			
<i>Callicebus calligatus</i>	<i>Callicebus caligatus</i>		
<i>Callicebus cinerascens</i>			
<i>Callicebus cupreus</i>			
<i>Callicebus donacophilus</i>			
<i>Callicebus dubius</i>			
<i>Callicebus hoffmannsi</i>			
<i>Callicebus modestus</i>			
<i>Callicebus moloch</i>		Y	
<i>Callicebus oenanthe</i>			
<i>Callicebus olallae</i>			
<i>Callicebus personatus</i>			
<i>Callicebus torquatus</i>			
<i>Callimico goeldii</i>		Y	
<i>Callithrix argentata</i>			
<i>Callithrix humeralifer</i>			
<i>Callithrix jacchus</i>		Y	Y

<i>Callithrix pygmaea</i>	<i>Cebuella pygmaea</i>	Y	Y
<i>Cebus albifrons</i>		Y	Y
<i>Cebus apella</i>		Y	Y
<i>Cebus capucinus</i>			Y
<i>Cebus olivaceus</i>			Y
<i>Cercocebus aterrimus</i>	<i>Lophocebus aterrimus</i>		
<i>Cercocebus galeritus</i>			Y
<i>Cercocebus torquatus</i>		Y	Y
<i>Cercopithecus ascanius</i>		Y	Y
<i>Cercopithecus campbelli</i>			
<i>Cercopithecus cephus</i>			
<i>Cercopithecus denti</i>	<i>Cercopithecus wolfi denti</i>		
<i>Cercopithecus diana</i>	<i>Cercopithecus roloway</i>		Y
<i>Cercopithecus dryas</i>			
<i>Cercopithecus erythrogaster</i>			
<i>Cercopithecus erythrotis</i>			
<i>Cercopithecus hamlyni</i>			
<i>Cercopithecus lhoesti</i>			
<i>Cercopithecus mitis</i>		Y	Y
<i>Cercopithecus mona</i>			Y
<i>Cercopithecus neglectus</i>			
<i>Cercopithecus nictitans</i>		Y	
<i>Cercopithecus petaurista</i>			
<i>Cercopithecus pogonias</i>			
<i>Cercopithecus preussi</i>			
<i>Cercopithecus salongo</i>			
<i>Cercopithecus solatus</i>			
<i>Cercopithecus wolfi</i>			
<i>Cheirogaleus major</i>		Y	
<i>Cheirogaleus medius</i>		Y	
<i>Chiropotes albinus</i>			
<i>Chiropotes satanas</i>			
<i>Chlorocebus aethiops</i>	<i>Cercopithecus aethiops</i>		Y
<i>Colobus angolensis</i>			
<i>Colobus guereza</i>			
<i>Colobus polykomos</i>			
<i>Colobus satanas</i>			
<i>Daubentonia madagascariensis</i>		Y	Y
<i>Erythrocebus patas</i>		Y	Y
<i>Eulemur coronatus</i>	<i>Petterus coronatus</i>		
<i>Eulemur fulvus</i>	<i>Petterus fulvus, Lemur fulvus</i>	Y	Y
<i>Eulemur macaco</i>	<i>Petterus macaco</i>		Y
<i>Eulemur mongoz</i>	<i>Petterus mongoz</i>	Y	
<i>Eulemur rubriventer</i>	<i>Petterus rubriventer</i>		
<i>Euoticus elegantulus</i>			
<i>Euoticus inustus</i>	<i>Galago matschiei</i>		
<i>Galago alleni</i>			
<i>Galago granti</i>	<i>Galago senegalensis granti</i>		
<i>Galago moholi</i>			
<i>Galago senegalensis</i>			
<i>Galagoides demidoff</i>	<i>Galago demidovii</i>		
<i>Galagoides zanzibaricus</i>			
<i>Gorilla gorilla</i>		Y	Y
<i>Hapalemur aureus</i>			
<i>Hapalemur griseus</i>			
<i>Hapalemur simus</i>			
<i>Hylobates agilis</i>	<i>H. lar agilis</i>		
<i>Hylobates concolor</i>	<i>Nomascus concolor</i>		Y
<i>Hylobates gabriellae</i>	<i>Nomascus gabriellae</i>		Y
<i>Hylobates hoolock</i>	<i>Bunopithecus hoolock</i>		

<i>Hylobates klossii</i>	<i>Hylobates klossi</i>		
<i>Hylobates lar</i>		Y	
<i>Hylobates leucogenys</i>	<i>Nomascus leucogenys</i>		
<i>Hylobates moloch</i>			
<i>Hylobates muelleri</i>	<i>H. agilis muelleri</i>		
<i>Hylobates pileatus</i>			Y
<i>Hylobates syndactylus</i>	<i>Symphalangus syndactylus</i>		
<i>Indri indri</i>			
<i>Lagothrix flavicauda</i>			
<i>Lagothrix lagotricha</i>	<i>Lagothrix lagotricha</i>		
<i>Lemur catta</i>		Y	Y
<i>Leontopithecus chrysomelas</i>			Y
<i>Leontopithecus chrysopygus</i>			
<i>Leontopithecus rosalia</i>			
<i>Lepilemur dorsalis</i>			
<i>Lepilemur edwardsi</i>			
<i>Lepilemur leucopus</i>			
<i>Lepilemur microdon</i>			
<i>Lepilemur mustelinus</i>			
<i>Lepilemur ruficaudatus</i>		Y	
<i>Lepilemur septentrionalis</i>			
<i>Lophocebus albigena</i>	<i>Cercocebus albigena</i>	Y	Y
<i>Loris tardigradus</i>		Y	Y
<i>Macaca arctoides</i>	<i>Macaca speciosa</i>		Y
<i>Macaca assamensis</i>			
<i>Macaca cyclopis</i>			
<i>Macaca fascicularis</i>	<i>Macaca irus, Macaca cynomolgus</i>		Y
<i>Macaca fuscata</i>			Y
<i>Macaca maura</i>	<i>Macaca maurus</i>		
<i>Macaca mulatta</i>		Y	Y
<i>Macaca nemestrina</i>			Y
<i>Macaca nigra</i>			
<i>Macaca ochreata</i>			
<i>Macaca radiata</i>			Y
<i>Macaca silenus</i>			Y
<i>Macaca sinica</i>			
<i>Macaca sylvanus</i>			Y
<i>Macaca thibetana</i>			Y
<i>Macaca tonkeana</i>	<i>Macaca tonkeana</i>		Y
<i>Mandrillus leucophaeus</i>	<i>Papio leucophaeus</i>		
<i>Mandrillus sphinx</i>	<i>Papio sphinx</i>	Y	Y
<i>Microcebus coquereli</i>	<i>Mirza coquereli</i>		
<i>Microcebus murinus</i>		Y	
<i>Microcebus rufus</i>			
<i>Miopithecus talapoin</i>	<i>Cercopithecus talapoin</i>	Y	
<i>Nasalis concolor</i>	<i>Simias concolor</i>		
<i>Nasalis larvatus</i>		Y	
<i>Nycticebus coucang</i>		Y	
<i>Nycticebus pygmaeus</i>			
<i>Otolemur crassicaudatus</i>	<i>Galago crassicaudatus</i>	Y	Y
<i>Otolemur garnettii</i>			Y
<i>Pan paniscus</i>		Y	Y
<i>Pan troglodytes</i>		Y	Y
<i>Papio anubis</i>	<i>Papio hamadryas anubis</i>	Y	Y
<i>Papio cynocephalus</i>	<i>Papio hamadryas cynocephalus</i>	Y	Y
<i>Papio hamadryas</i>	<i>Papio hamadryas hamadryas</i>	Y	Y
<i>Papio papio</i>	<i>Papio hamadryas papio</i>		Y
<i>Papio ursinus</i>	<i>Papio hamadryas ursinus</i>		Y
<i>Perodicticus potto</i>		Y	
<i>Phaner furcifer</i>			

<i>Pithecia aequatorialis</i>			
<i>Pithecia albicans</i>			
<i>Pithecia irrorata</i>			
<i>Pithecia monachus</i>	<i>Pithecia monacha</i>	Y	
<i>Pithecia pithecia</i>			
<i>Pongo pygmaeus</i>		Y	Y
<i>Presbytis comata</i>	<i>Presbytis aygula</i>		
<i>Presbytis frontata</i>			
<i>Presbytis geei</i>	<i>Trachypithecus geei</i> ; <i>Semnopithecus geei</i>		
<i>Presbytis melalophos</i>			
<i>Presbytis potenziani</i>			
<i>Presbytis rubicunda</i>			
<i>Presbytis vetulus</i>	<i>Trachypithecus vetulus</i> ; <i>Kasi vetulus</i> ; <i>Presbytis senex</i>		
<i>Procolobus badius</i>	<i>Colobus badius</i>	Y	Y
<i>Procolobus kirkii</i>	<i>Colobus kirkii</i>		Y
<i>Procolobus verus</i>			
<i>Propithecus diadema</i>			
<i>Propithecus tattersalli</i>			
<i>Propithecus verreauxi</i>		Y	Y
<i>Pygathrix avunculus</i>	<i>Rhinopithecus avunculus</i>		
<i>Pygathrix brelichi</i>	<i>Rhinopithecus brelichi</i>		
<i>Pygathrix nemaesus</i>		Y	
<i>Pygathrix roxellana</i>	<i>Rhinopithecus roxellana</i>		
<i>Saguinus bicolor</i>			
<i>Saguinus fuscicollis</i>			Y
<i>Saguinus imperator</i>			
<i>Saguinus inustus</i>			
<i>Saguinus labiatus</i>			Y
<i>Saguinus leucopus</i>			
<i>Saguinus midas</i>		Y	
<i>Saguinus mystax</i>			Y
<i>Saguinus nigricollis</i>			
<i>Saguinus oedipus</i>		Y	
<i>Saguinus tripartitus</i>			
<i>Saimiri boliviensis</i>			
<i>Saimiri oerstedii</i>			Y
<i>Saimiri sciureus</i>			Y
<i>Saimiri ustus</i>			
<i>Saimiri vanzolinii</i>			
<i>Semnopithecus entellus</i>	<i>Presbytis entellus</i>	Y	Y
<i>Tarsius bancanus</i>			
<i>Tarsius diana</i>			
<i>Tarsius pumilis</i>			
<i>Tarsius spectrum</i>			
<i>Tarsius syrichta</i>		Y	
<i>Theropithecus gelada</i>			Y
<i>Trachypithecus auratus</i>	<i>Presbytis aurata</i> , <i>P. auratus</i> ; <i>Trachypithecus aurata</i> ; <i>T. auratus</i> ; <i>Semnopithecus auratus</i> ; <i>S. aurata</i>		
<i>Trachypithecus cristatus</i>	<i>Presbytis cristata</i>		
<i>Trachypithecus francoisi</i>	<i>Presbytis francoisi</i> ; <i>Semnopithecus francoisi</i>		
<i>Trachypithecus johnii</i>	<i>Presbytis johnii</i> ; <i>Semnopithecus johnii</i> ; <i>Kasi johnii</i>		Y
<i>Trachypithecus obscurus</i>	<i>Presbytis obscura</i>		
<i>Trachypithecus phayrei</i>	<i>Presbytis phayrei</i>		
<i>Trachypithecus pileatus</i>	<i>Presbytis pileatus</i>		
<i>Varecia variegata</i>		Y	

211 **2. Brain volume measures**

212 Any study of primate brain evolution should note that only a small proportion of primate
213 brains have been measured, correlational analyses are employed, and conclusions must
214 be necessarily tentative. While information on neuronal and nonneuronal cell
215 composition would be extremely valuable for these comparative analyses, it is currently
216 not available for a sufficient number of primate species to conduct robust analyses [48-
217 50]. The most appropriate measure of brain size is controversial [1,50-52], so we
218 consider four popular brain volume measures:

219 (a) *neocortex ratio* (neocortex over rest of brain), chosen because it is the most
220 commonly used measure [11,53];

221 (b) *executive brain ratio* (neocortex and striatum over brainstem), supported by the
222 observation that the neocortex and striatum are functionally and phylogenetically linked
223 [54]. The widely used approach of using brainstem volume (sum of mesencephalon and
224 medulla oblongata) as a reference variable makes the assumption that such areas are
225 evolutionarily conservative [51,54]. This is preferable to using body mass as a reference
226 variable, since body mass, unlike brainstem volume, is a rather inaccurate measure of
227 body size [51];

228 (c) *neocortex size*, argued to be reflective of raw information processing power [55];

229 (d) *residuals of neocortex on rest of brain* (excluding cerebellum), used by [8]. This
230 measure avoids the risk of confounding enlarged neocortex volumes with diminished
231 cerebellum volumes, which may be important if the cerebellum plays a greater role in
232 cognition than previously thought, as suggested by Barton [56]. Rest of brain was used as
233 the reference variable here since brainstem data were not available for specimens
234 measured by MRI.

235 Residual measures (such as measure d) require a reference group to be defined
236 (here, all primates with available data), and species values will change with changes in
237 the reference group. Ratio and absolute brain component measures (measures a-c) carry
238 the advantage that they do not change with the reference group, but the disadvantage that
239 they correlate with body mass [1].

240

241 **3. Phylogenetic analyses**

242 Where we address evolutionary questions we correct for phylogeny using independent
243 contrasts implemented with the CAIC computer program [57]. CAIC makes fewer type I
244 errors than across-species analyses even when the branch lengths and phylogeny are
245 uncertain or inaccurate [57]. All brain and body sizes, apart from the executive brain
246 ratio, were natural-log transformed before taking contrasts since CAIC assumes that
247 different lineages are equally likely to make the same proportional change in size.
248 Across-species analyses were also conducted using these natural-log transformed values.
249 Independent contrasts were regressed through the origin using least-squares regression
250 [57]. The primate phylogeny was a composite tree [45], with a prior version of this tree
251 used for sensitivity analyses [47]. For the purposes of this analysis an assumption of
252 equal distances between phylogenetic nodes was made. We report the results of both
253 independent contrasts and species-level analyses because questions have been raised over
254 whether independent contrast analyses are always more valid than analyses treating
255 species as independent data points (e.g. [58]).

256

257 In order to investigate the distribution of primate g across the primate phylogeny, we
258 pooled data across species into genera and conducted a genus-level PCA, generating

259 primate g scores for each genus. These scores were then \ln transformed and converted to
260 z scores before using MacClade 4.08 [59] to reconstruct the evolution of primate g_{S1} . A
261 linear parsimony algorithm was applied with minimum values at each branch displayed
262 on figure 1b. We used the Purvis [45,47] phylogeny, despite the fact that a newer
263 phylogeny including 30 additional species had recently become available [60]. That is
264 because the Purvis phylogeny adopts nomenclature closer to that of the authors of the
265 original articles that are the sources of our data, easing unambiguous allocation of a
266 report of behavioural plasticity and phylogenetic analysis. Reconstructing the evolution
267 of primate g using the new phylogeny gives identical results to those presented.

268

269 The reconstruction process was repeated at the species level, generating qualitatively
270 similar results (i.e. evidence for convergent evolution favouring high intelligence in
271 Hominoidea, *Macaca*, *Cebus*, and *Papio*). In the species-level analysis we addressed the
272 concern that the data for little-studied species may be unreliable, by repeating this
273 procedure removing species with a research effort of less than 10, <20 and <40 studies in
274 the database. Again, the qualitative picture did not change, with evidence for independent
275 evolutionary events favouring high intelligence in Hominoidea, *Macaca*, *Cebus*, and
276 possibly *Papio*. We are less confident of *Papio*, where the high score reflects a relatively
277 small number of reports of our measures of behavioural flexibility in comparatively little
278 studied species (*Papio papio*, *Papio ursinus*). The analyses also hint at selection for
279 intelligence in other lineages (e.g. *Saguinus*, *Erythrocebus*), but these trends require
280 further data for confirmation.

281

282

282 **4. Confounding variables and reliability**

283 Inter-observer reliabilities in data categorization are high (0.83-0.95) [3]. We conducted
284 analyses to statistically control for common ancestry, brain size, body mass, research
285 effort, or correlated error in research effort, and at the genus level (see main text or
286 below). The prominence of a single general intelligence component in a PCA (or factor
287 in an FA) holds in all cases. Confidence in the methods is enhanced by the observation
288 that other studies using similar procedures have investigated but not found evidence for
289 various forms of bias. For instance, Lefebvre *et al.* [4,5,61] collected avian foraging
290 innovation data from published literature, and demonstrated that their findings were not
291 affected by population size, journal source, editorial policy, research effort, or observer
292 bias (ornithological interest measured as the frequency of photographs in birding
293 publications and by questionnaire). Furthermore, Lefebvre's innovation measure also
294 correlates with laboratory measures of learning performance [62]. In primates, previous
295 work examining pairwise correlations between innovation, social learning, and tool use
296 found that the results were unaffected by the removal of reports from captivity or where
297 human intervention was implicated [3].

298

299 While we cannot exclude the possibility that some of our measures, particularly social
300 learning and tactical deception (where underlying processes are difficult to assess
301 without experimentation, see e.g. [63], may be prone to reporting biases beyond those
302 associated with research effort [3,64]. Such reporting biases could result from
303 assumptions about the cognitive abilities of primates closely related to humans, or from
304 differences in cognitive performance in the laboratory priming researchers to look for
305 and report behavioural flexibility in high-performing species. We endeavoured to

306 establish whether such biases may account for our major findings. For instance, if
307 primatologists were prone to expect more intelligent behaviour amongst great apes than
308 other primates, and if such expectations led to inadvertent reporting biases, our findings
309 should be substantially weakened by repeating the analyses with the great apes removed.
310 Such an analysis would also control for the possibility that the observed patterns of
311 variation are solely reliant on the great apes. In fact, removing the great apes does not
312 affect our findings (see below).

313

314 Combined, these observations, and the positive associations of primate g with brain size
315 and laboratory performance (see main text), suggest that our methods generate a robust
316 operational measure of behavioural flexibility, or intelligence. Although the tactical
317 deception database [15] has received criticism (see peer commentary on Whiten & Byrne
318 [16]), it forms only a small part of our data and its exclusion does not change our basic
319 findings. More generally, questions over the validity of any single measure do not negate
320 the message that multiple cognitive measures covary (Table 3).

321

322 **5. Principal components and factor analyses**

323 We conducted principal components analyses, utilizing 5-8 measures of cognitive
324 performance, employing a roots-greater-than-1 extraction criterion, all of which revealed
325 a single dominant component that we call ‘primate g_s ’. Where more than one component
326 was extracted (8-variable analysis), axes were rotated to maximize the variance
327 accounted for, but retained an orthogonal (independent) component design so that the
328 axes can be regarded as powerful descriptive dimensions. We also conducted factor
329 analyses to investigate whether our findings were robust to the analysis technique.

330

331 *(a) Extant species variation*

332 The reported observation of a single dominant component in a PCA (and a single
333 dominant factor in FA) of measures of cognitive ability is not an artefact of the
334 covariance of each individual measure with brain volume, body mass or error variance in
335 research effort, nor does it result from our removal of cases that qualify for more than
336 one variable. Equivalent results were found for PCA using residuals of each cognitive
337 measure from multiple regressions that include (1) relative brain volume ($\chi^2 = 82.97, p <$
338 0.0001 , variance contribution = 84%), (2) body mass ($\chi^2 = 135.40, p < 0.0001$, variance
339 contribution = 59%), (3) employing five independent measures of research effort ($\chi^2 =$
340 $107.32, p < 0.0001$, variance contribution = 81%), (4) without accounting for research
341 effort ($\chi^2 = 189.00, p < 0.0001$, variance contribution = 72%), (5) without removal of
342 cases that simultaneously qualified for more than one measure of behavioural flexibility
343 ($N = 69, \chi^2 = 273.68, p < 0.0001$, variance contribution = 74%), and (6) with the great
344 apes removed ($\chi^2 = 42.04, p < 0.0001$, variance contribution = 45%).

345

346 *(b) Independent contrast variation*

347 Independent contrasts were forced through the origin by duplicating each data row and
348 taking the reciprocal of the duplicated data [65]. (See [66] for an alternative procedure).
349 Since this procedure inflates χ^2 values [67], we report significance values from analyses
350 not forced through the origin, which produced similar results.

351

352

352 **Supplemental References**

353

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